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Interaction of C/EBP-beta and NF-Y factors constrains activity levels of the nutritionally controlled promoter IA expressing the acetyl-CoA carboxylase-alpha gene in cattle

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Abstract

Background: The enzyme acetyl-CoA carboxylase-alpha (ACC- α) is rate limiting for de novo fatty acid synthesis. Among the four promoters expressing the bovine gene, promoter IA (PIA) is dominantly active in lipogenic tissues. This promoter is in principal repressed but activated under favorable nutritional conditions. Previous analyses already coarsely delineated the repressive elements on the distal promoter but did not resolve the molecular nature of the repressor. Knowledge about the molecular functioning of this repressor is fundamental to understanding the nutrition mediated regulation of PIA activity. We analyzed here the molecular mechanism calibrating PIA activity.

Results: We finely mapped the repressor binding sites in reporter gene assays and demonstrate together with Electrophoretic Mobility Shift Assays that nuclear factor-Y (NF-Y) and CCAAT/enhancer binding protein- β (C/EBP β) each separately repress PIA activity by binding to their cognate low affinity sites, located on distal elements of the promoter. Simultaneous binding of both factors results in strongest repression. Paradoxically, over expression of NFY factors, but also - and even more so - of C/EBP β significantly activated the promoter when bound to high affinity sites on the proximal promoter. However, co-transfection experiments revealed that NF-Y may eventually diminish the strong stimulatory effect of C/EBP β at the proximal PIA in a dose dependent fashion. We validated by chromatin immunoprecipitation, that NF-Y and C/EBP factors may physically interact.

Conclusion: The proximal promoter segment of PIA appears to be principally in an active state, since even minute concentrations of both, NF-Y and C/EBP β factors can saturate the high affinity activator sites. Higher factor concentrations will saturate the low affinity repressive sites on the distal promoter resulting in reduced and calibrated promoter activity. Based on measurements of the mRNA concentrations of those factors in different tissues we propose that the interplay of both factors may set tissue-specific limits for PIA activity.

Keywords: ACC-alpha, *Bos taurus*, CCAAT-enhancer binding protein, Fat synthesis, Gene regulation, Nuclear factor Y

Background

Acetyl-CoA carboxylase-alpha (ACC- α) is the rate-limiting enzyme in de novo synthesis of long-chain fatty acids [1]. It thus plays an important role in controlling lipid metabolism [2]. Dysregulation of lipid metabolism in human and mouse is known to result in metabolic diseases, such as obesity and diabetes, and may also

accompany cancer [3,4]. It may also result in severe metabolic disorders in lactating cows [5]. Hence, an improved understanding of the regulation of ACC- α activity might help developing new strategies to avoid health problems associated with perturbed fat metabolism in keeping of dairy cattle.

Multiple promoters express the mammalian ACC- α encoding-gene. Their activities are differentially influenced by diets and hormones (e.g. insulin and glucagon) [1,6]. The bovine gene was previously known to be expressed by three promoters [7-9]. Among them, the

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5'-most located promoter was designated as "PI" and was supposed to be nutritionally regulated. However, we recently identified an additional fourth promoter located ~41 kbp more upstream, similar as previously reported from sheep [10]. We validated in separate experiments that this promoter is also active in cattle, mainly in the brain (EMBL Acc No FN185962, FN185963). To avoid confusion regarding the designation of the various promoters, we adopt here the recently proposed nomenclature of ACC- α promoters (Additional file 1: Figure S1) [6]. Thus, we now name, for cattle the most 5'-located promoter as PI and designate the nutritionally regulated promoter as PIA (previously known as "PI" [7]). We keep the previous designations for the constitutive promoter PII [8] and the mammary gland expressed and lactationally activated PIII [9].

PIA (Additional file 1: Figure S1), but not the other promoters, is known as the dominant ACC- α promoter of lipogenic tissues, such as liver, adipose tissues and mammary gland in ruminant and rodent species [11-13]. Activity of this promoter is controlled by insulin through an element binding the upstream stimulatory factors (USF1 or USF2) [14]. Glucose regulates PIA via a carbohydrate response element (ChoRE) located on the proximal promoter [15].

Promoter PIA is basically repressed [7,16,17]. Coarse mapping of structural elements necessary for repression of PIA in cattle had revealed that the repressor consists of potentially three distinct elements which are separated by more than 1000 bp [7]. The molecular nature of the repressor and the repressive mechanism controlling PIA activity remained unknown, so far. Cooperation of at least two of these structural elements was shown to be necessary in order to exert repression [7]. PIA from rat is also repressed by a distal element [16]. Over-expression of C/EBP α or - β factors may overcome repression of the rat PIA by binding to the proximal promoter and stimulate its activity in reporter gene assays [16,17].

C/EBP factors are ubiquitously expressed and are known to play an essential role in controlling the expression of genes with relevance for energy metabolism [18,19]. The C/EBP factor family includes six structurally related members (α , β , δ , ϵ , γ , ζ). Their specific N-termini encompass not only activation domains but also repression domains. The conserved C-termini constitute a DNA binding domain while an internal leucine zipper domain is necessary for factor dimerization [20]. These factors have a quite complex repertoire of mechanisms to eventually regulate expression of their target genes, involving factor dimerization, either as homo- or hetero-mer. Differential phosphorylation and synthesis of different isoforms were also shown to regulate activity of those factors [20,21].

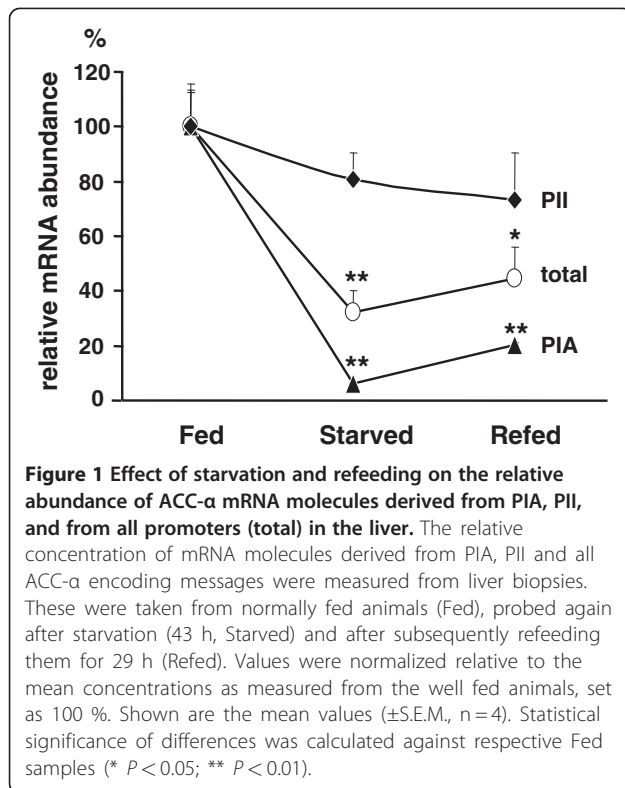
C/EBP α is known to interact with NF-Y factors [22,23]. The ubiquitously expressed NF-Y factor family consists of three subunits, NF-YA, -B and -C. Subunits -B and -C are constitutively expressed, while the expression of NF-YA is regulated [24,25]. NF-YB and -C have to form a dimer before NF-YA can bind to the complex. Only that trimer of NF-Y factors can bind to a CCAAT-box on the DNA. Mutation of three amino acids in the binding domain of the NF-YA subunit abrogates DNA-binding of the entire NF-Y complex [26]. NF-Y may act as a bifunctional regulator by either repressing or activating transcription during different physiological conditions [27].

It was found in rat and goat that the presence of this distal repressor reduces the insulin mediated stimulation of PIA activity [14,15]. Hence, quite obviously does the activity of the repressor influence and eventually bias the analysis of the nutritionally and hormone mediated fine tuning of PIA activity. Therefore, we set out to characterize the molecular nature of that general repressor of PIA in cattle. We first finely mapped the cis-relevant repressive elements and determined in mutation analyses and EMSA assays that NF-Y and C/EBP factors act as repressors. Surprisingly, we found that these factors both are also crucial enhancers of PIA activity, if they are bound to the proximal promoter. We provide evidence that interaction in trans allows the distally bound repressors to inhibit the stimulatory effect of the proximally bound partner factors. Measurements of the endogenous mRNA levels suggest that the proportion of these factors may set coarse tissue-specific limits for the overall PIA promoter activity.

Results

PIA is the nutritionally regulated promoter

We validated first that indeed the activity of PIA is nutritionally controlled, using the nutritionally modulated expression of the ACC- α in liver as paradigm. To this end we selected four first lactating cows and took biopsies from their livers. Subsequently the cows were starved for 43 h, and the livers were biopsied again. Sampling was repeated subsequent to refeeding the animals for 29 h again. Starvation decreased the total ACC- α mRNA content down to ~32% of that found in the livers of the well fed animals (Figure 1). This decrease was caused primarily by a cessation of PIA activity, as indicated by a >94% reduction of PIA derived transcripts. The transcripts derived from the house keeping promoter (PII), on the other hand, only decreased by ~19%. The data show that the level of PIA derived transcripts in the liver reflects the metabolic state of the animal, better than the concentration of all ACC- α -encoding messages. Also,



gross alterations in PIA activity influence the total amount of ACC-α transcripts. Strong nutrition-dependent regulation of PIA activity in the liver was also recorded in a different experiment, in which a group of six cows was kept on a minimal energy, straw based diet for 60 h [28]. This diet supplied only 16 % of the energy contained in the normal diet having been fed to the control group. The concentration of PIA derived ACC-α transcripts was proportionally reduced, down to 19 % (Table 1).

Distal NF-Y and C/EBP binding sites are involved in PIA repression

We finely mapped with serial deletions of promoter segments the structural elements contributing to PIA repression in murine mammary epithelial model cells (MEC) HC-11. MEC cells are a relevant model for fatty acid synthesis, since they synthesize in vivo abundant amounts of milk fat [29] and the previous functional characterization of PIA had in part been conducted using these cells [7]. We found that two elements (A and C) from among three potentially relevant elements must be present to repress the PIA activity (Figure 2A and Additional file 1: Figure S2A). Deletion of either element increased the promoter activity > 3-fold. Strongest promoter activation was achieved, if both repressive sites were deleted. We cloned to this end a short segment comprising only 127 bp of the proximal PIA promoter (Driver, Figure 2A and Additional file 1: Figure S2A). This element very actively expresses the reporter gene, resulting in approximately 10-fold of the reporter activity measured from the promoter-less pGL3b (Figure 3B), confirming previous observations [7]. The extent of enhanced activity of this short minimal promoter indicates an additive effect of both repressive sites.

More detailed deletion analyses revealed that the 5'-located half of element A is the key component of the repressive element (Figure 2A and Additional file 1: Figure S2B). Abolishing the NF-Y factor binding site in this element via point mutation increased promoter activity ~3-fold, indicating some relief of repression (Figure 2B, clone Am). EMSA assays demonstrated that NF-Y factors can indeed bind to a probe harboring the 5'-half of element A (Figure 2C, lane4). We used for these assays nuclear extracts from murine HC-11 cells, from primary bovine MEC cells (pbMEC) or from the lactating udder of a cow (bMG). The specific complex (NF-Y) could be inhibited by the addition of a 100-fold molar excess of a competitor, harboring a NF-Y consensus motif (Figure 2C, lane3) embedded into otherwise completely PIA-

Table 1 Relative mRNA levels of ACC-α PIA, NF-YA and C/EBPβ in various tissues under different nutritional conditions

Diet	Copies	Liver		Adipose		Mammary gland	
		Straw	Normal	Fat	Starch	Starch	Fat
ACC-α PIA	$\times 10^4$	0.10 ± 0.01^a	0.52 ± 0.22^b	6.81 ± 2.24	8.49 ± 4.86	19.79 ± 4.79	42.85 ± 12.98
NF-YA	$\times 10^3$	8.94 ± 1.05	7.64 ± 0.71	10.41 ± 0.97	18.57 ± 7.61	8.38 ± 1.48	7.56 ± 1.06
C/EBPβ	$\times 10^6$	4.37 ± 0.25	3.71 ± 0.80	2.45 ± 0.35	3.08 ± 1.10	0.62 ± 0.09^a	0.36 ± 0.06^b
NFYA:C/EBPβ	$\times 10^{-3}$	2.06 ± 0.24	2.44 ± 0.64	4.69 ± 0.71	6.45 ± 0.90	14.07 ± 1.87^a	25.89 ± 5.08^b
NEL	MJ	18.0 ± 1.1	110.2 ± 11.2	nd	nd	nd	nd
Fat	g/kg DM	nd	nd	51	28	28	51
Carbohydrate	g/kg DM	nd	nd	31	100	100	31
n	6	6	8	8	8	8	8

Mean values (\pm S.E.M.). Different superscripts ("a" and "b") indicate statistically different values ($P < 0.05$) within the respective tissue. NEL, Net Energy content for Lactation; nd, not determined; n, number of animal.

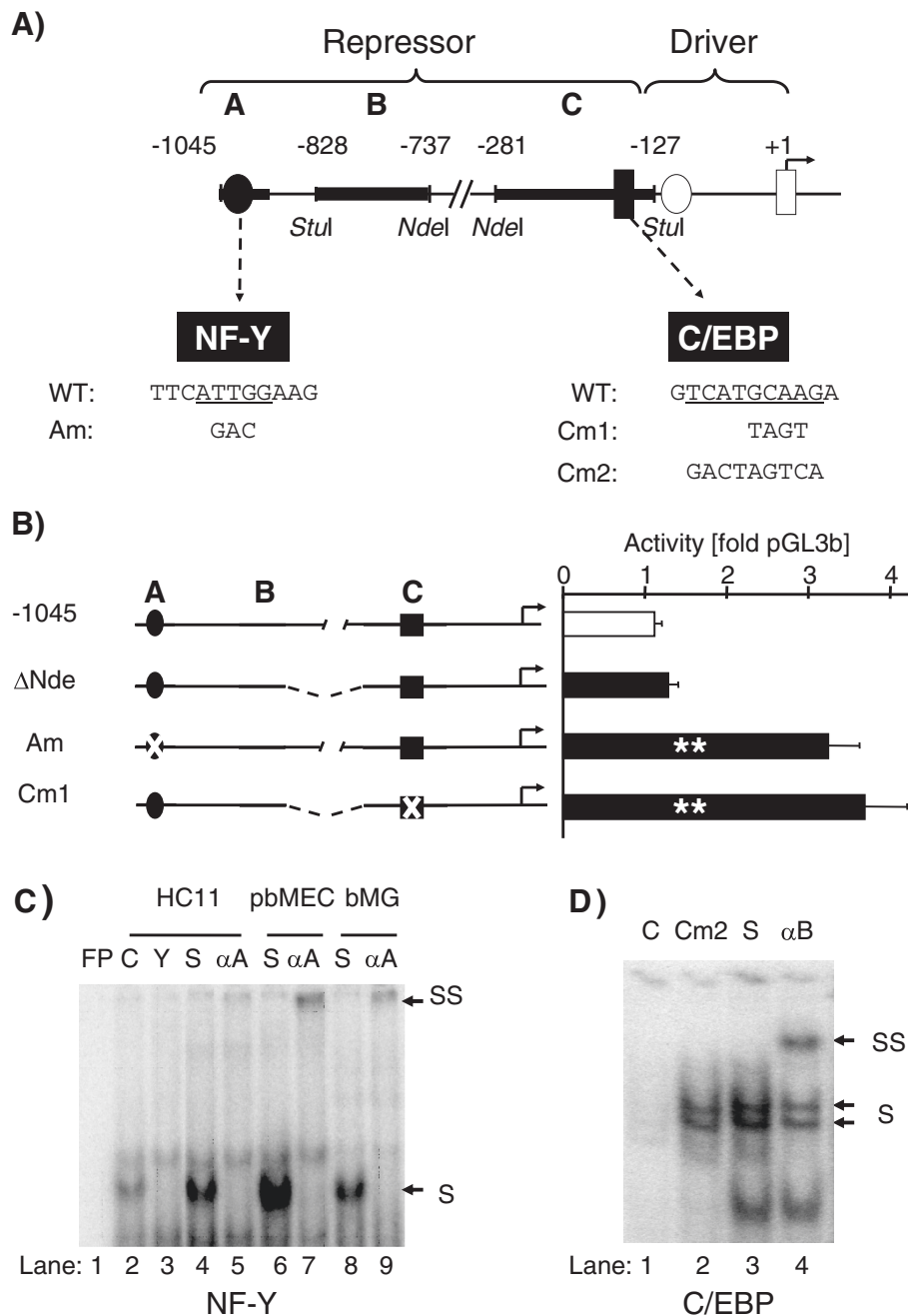


Figure 2 NF-Y and C/EBP binding sites in element A and C respectively in the distal region are involved in PIA repression. (A) Genomic organization of PIA. The distal region includes three putative repressive elements, as revealed by coarse mapping analyses involving *Stul* and *NdeI* deletions, respectively (filled boxes, element A, B, C). Element A harbors a NF-Y attachment site (filled circle), and element C contains a C/EBP binding site (filled square). The core binding sequences of those factors and mutational derivatives are shown. The proximal driving region features a NF-Y site (open circle) and a C/EBP binding site (open square). '+1' represents the tsp. (B) Abrogation of NF-Y binding and C/EBP binding in the distal PIA relieves repression in reporter gene assay. Shown are mean values (+ S.E.M.) of at least three independent experiments, each assayed in triplicate. Significance of difference relative to construct -1045 (WT) is indicated (** $P < 0.01$). (C) EMSA analysis of transcription factors binding to element A. The labeled probe A_WT (Additional file 1: Table S4) is bound by NF-Y factors from nuclear extracts of HC-11, pbMEC and bMG ("S") and is supershifted (SS) by the addition of antibodies against NF-YA (αA). Lanes: FP, free probe; C and Y, competition with 100-fold molar excess of the unlabeled A_WT probe and a probe harboring an NF-Y consensus site respectively (Additional file 1: Table S4). (D) EMSA validation that C/EBP factors bind to element C. Nuclear extracts from HC-11 cells were incubated with the radioactively labeled probe C_WT (Additional file 1: Table S4). Lane: C, competition with 100-fold molar excess of unlabeled C_WT; S, shift; αB, supershift ("SS") with anti-C/EBP antibodies; Cm2, competition with 100-fold molar excess of the unlabeled probe mutated as shown in Figure 2A.

(See figure on previous page.)

Figure 3 Binding of NF-Y and C/EBP factors to the proximal PIA drives activity. (A) Map and sequences of the NF-Y and C/EBP binding sites on the proximal PIA. Core motifs (underlined) of the respective binding sequences and mutations are shown, as they have been included in EMSA and reporter gene analyses. The tsp in the C/EBP motif ("A" nucleotide, bold, italicized) is indicated. (B) Mutation of the binding sites of NF-Y (Ym) as well as of C/EBP (m1, m2) significantly reduce the reporter genes activity (open columns). Joint co-transfection of expression constructs for all three NF-Y factors (NF-YA -B, -C) significantly increase activity of WT-promoter, but not of the construct Ym. Mean values (\pm S.E. M.) from one out of three experiments (Ym) or six independent experiments (C/EBP mutation), always assayed in triplicate and normalized against the WT construct. (C) EMSA analyses of the proximal NF-Y and C/EBP binding sites. Left: Nuclear extracts from HEK293 cells over-expressing jointly the factors NF-YA, -B, -C were incubated with the probe harboring the proximal NF-Y binding site (pNF_WT, Additional file 1: Table S4). The shifted band (S) was supershifted (SS) by anti-NF-YA antibodies (α A). Right: The labeled probe was the proximal C/EBP site (pCE_WT, Additional file 1: Table S4). Multiple specific shift bands (S) appeared with nuclear extracts from HC-11 cells, pbMEC, or bMG. Anti-C/EBP antibodies (α B) caused supershifts (SS). Lane designations are similar to Figure 2C and D; m1, competition with 100-fold molar excess of unlabeled probe, mutated as shown in Figure 3A. "ns" represents a non-specific band.

Additional file 1: Figure S2D). Using this candidate area as a probe in EMSA analyses showed that mutation of the C/EBP binding site abolished factor binding (Figure 2D, lane2, Cm2) and an antibody against C/EBP β supershifted this band (Figure 2D, lane4). Mutation of the C/EBP-core recognition motif relieved repression to the same extent as deleting that element (compare Figure 2B, clone Cm1 vs Additional file 1: Figure S2A, clone Δ Stu). Hence, the relevant DNA-sequence element within the repressive element C is a C/EBP binding site.

Our data together show that the structure of the bovine PIA repressor consists of a distal NF-Y binding site and a more proximally residing C/EBP binding site (Figure 2A). These sites cooperate functionally to exert full repression.

C/EBP and NF-Y factors activate PIA by binding to proximal promoter segments

Unexpectedly, cloning and co-transfecting together expression constructs for all three different NF-Y factors (A, B, C) consistently increased the activity of our long PIA reporter construct. Moreover, we previously reported that cotransfecting expression constructs for any of the C/EBP-factors - α , - β , - δ , - ϵ increased significantly the activity of this promoter. C/EBP β had the strongest activation potential [11]. Sequence analysis of the PIA DNA sequence revealed putative binding sites for both factors, indeed, located on the proximal promoter (Figure 3A). These are located on our minimal-127 bp promoter construct. When expression constructs for all three NF-Y subunits (A, B, C) were jointly co-transfected together with the reporter gene, they increased the reporter activity by 60–100% (Figure 3B). Abrogation of the NF-Y binding site via point mutation reduced the promoter activity by \sim 60% (Figure 3B, clone Ym). In support, co-transfected NF-Y expression constructs did not enhance the activity of the reporter harboring the mutated NF-Y binding site (Figure 3B, clone Ym, black column). EMSA analysis confirmed that over-expressed NF-Y-factors indeed may bind to this

element (Figure 3C, lane4). The unlabeled probe bearing a NF-Y consensus binding site (Y) competed away the shift band (Figure 3C, lane3), and anti-NF-YA antibodies (α A) supershifted this band (Figure 3C, lane5).

EMSA assays demonstrated that nuclear extracts from HC-11 clearly shifted a radioactively labeled probe bearing the proximal C/EBP motif (Figure 3C, lane9) which could also be supershifted by antibodies specific for C/EBP β (Figure 3C, lane10, 12, 14). Similarly, nuclear extracts derived either from pbMEC or bMG also formed several complexes with that probe, some of which could be supershifted by those antibodies. Abrogating this site via point mutation reduced the promoter activity down to \sim 30% of the WT activity (Figure 3B, clones m1 and m2). Considering that this C/EBP binding site comprises the most 3'-located transcriptional start point (tsp) of PIA [7], we inserted two different mutations into this sequence element, one eliminating the "A" nucleotide forming the WT tsp while the other maintained the tsp (Figure 3B; clones m1 and m2, respectively). Both mutations resulted in a comparable and diminished activity of the reporter. The data together show that C/EBP as well as NF-Y factors activate the proximal PIA promoter.

The distal sites bind the respective factors with much lower affinities than the proximal sites

The data presented the paradox that NF-Y as well as C/EBP-factors may either function as repressor or as driver of PIA activity, depending on binding to either the distal or the proximal sites on the promoter. We realized, however, in the course of the EMSA analyses that the binding affinities of the distal and the proximal sites were different. To quantitatively evaluate the differential binding affinities of the distal and proximal DNA-sequence motifs for both, NF-Y and C/EBP β , we labeled as probes double stranded oligonucleotides harboring the core consensus motifs for either factor and embedded these into DNA-sequences which were

otherwise entirely unrelated to the ACC- α PIA (NF-Y consensus, C/EBP consensus oligonucleotides, Additional file 1: Table S4). However, we choose as unlabeled competitors double stranded oligonucleotides with sequences representing either the distal or the proximal binding sites for the respective factors (for NF-Y: oligonucleotides A_WT, and pNF_WT; for C/EBP: oligonucleotides C_WT, pNF_WT, respectively; Additional file 1: Table S4). We used nuclear extracts for the assays prepared from cells overexpressing either all three NF-Y factors, or the factor C/EBP β . Regarding the proximal NF-Y binding site (Figure 4A), we found that a 25-fold molar excess of the competitor would reduce the amount of the shifted probe down to 20%

(Figure 4A, lane 5 and Additional file 1: Figure S3). In stark contrast, it required as much as a 200-fold molar excess of the distal attachment site to obtain the same degree of competition for the probe (Figure 4A, lane 13 and Additional file 1: Figure S3). An even larger difference in competition efficiency was observed for the C/EBP attachment sites (Figure 4B). As little as a 10-fold molar excess of the proximal site competitor reduced the shifted amount down to less than 20% (Figure 4B, lane 4 and Additional file 1: Figure S3). However, even a 200-fold molar excess of the distal site would still leave behind ~40% of the radioactive probe in the shifted band (Figure 4B, lane 13 and Additional file 1: Figure S3). These data demonstrate that the

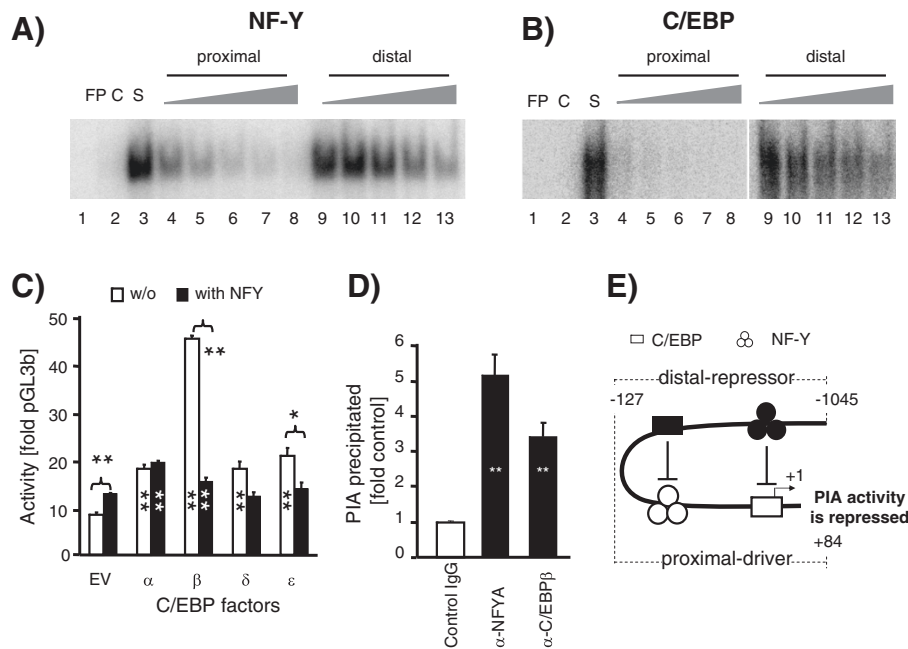


Figure 4 Comparison of the binding affinities of the distal and proximal NF-Y and C/EBP sites and influence of these factors on the PIA activity. (A) Affinity comparison of the proximal and distal NF-Y binding sites of PIA. Nuclear extracts were prepared from HEK293 cells jointly over-expressing the three bovine NF-Y factors (A, B, C). Sequences of the labeled NF-Y consensus probe and of the unlabeled competitor of the distal NF-Y (A_WT) and proximal NF-Y sites (pNF_WT) are shown in Additional file 1: Table S4 *cf* (Figures 2A and Figure 3A). Lane: FP, free probe; C, competition with 100-fold molar excess of unlabeled probe (self); S, shift. The fold molar excess of the respective unlabeled competitors added to the reactions, as resolved in lanes 4–8 or lanes 9–13 were 10, 25, 50, 100, 200-fold of the labeled probe. (B) Same as A, but evaluating the C/EBP binding sites. These samples were run out on the same gel. Nuclear extracts were prepared from HC-11 cells over-expressing the DNA binding domain of the bovine C/EBP β factor. Sequences of the labeled C/EBP consensus probe, the unlabeled oligonucleotide competitors of C/EBP distal (C_WT) and C/EBP proximal (pCE_WT) are shown in Additional file 1: Table S4 *cf* (Figure 2A and Figure 3A). (C) Effects of NF-Y and C/EBP factors on the proximal PIA activity. The proximal PIA reporter construct –127 was transfected into HC-11 cells, together with the empty cloning vector (EV) or with constructs expressing C/EBP β or NF-Y subunits, as indicated. The functional NF-Y trimer can quench this effect. Results (\pm S.E.M.) are representative for two experiments, each assayed in triplicate. Statistical significance of the differences was calculated relative to the control having been cotransfected only with empty vector (grey column), $**P < 0.01$. (D) NF-Y factors interact with C/EBP β having bound to PIA. HEK293 cells were co-transfected with the –127 PIA reporter gene and expression constructs for the factors C/EBP β (tagged with a flag), NF-YAm29, -YB, -YC. Nuclei were used for ChIP assays using antibodies against C/EBP β or NF-YA or control IgG. Data are mean values (\pm S.E.M.) of four qPCR assays of the amount of recovered PIA promoter molecules ($**P < 0.01$, as compared to control IgG). (E) A master control unit modulates PIA activity. NF-Y and C/EBP factors (black) having bound to the distal repressive factor binding sites may be looped back to physically contact the respective other factor (open) having bound to the proximal driver region. This interaction lowers PIA activity. The map is not drawn to scale. “+1” represents the tsp.

distal attachment sites have much lower binding affinities for the respective factors than those proximal sites.

NF-Y quenches the stimulating effect of C/EBP β

We analyzed how NF-Y and C/EBP β together might repress PIA. Therefore, we co-transfected both factors either alone or jointly together and recorded the effect on the activity of the short PIA promoter segment. Both factors could separately activate the PIA reporter gene activity (Figure 4C and Additional file 1: Figure S4B). Co-transfection of expression constructs for all three NF-Y factors abolished the stimulatory effect of C/EBP β (Figure 4C, Additional file 1: Figure S4A and Additional file 1: Figure S4B). Quenching of the C/EBP β associated transactivation potential is dosage-dependent, since increasing amounts of transfected DNA caused stronger quenching of the stimulatory capacity of C/EBP β (Additional file 1: Figure S4A). Quenching required that all three NF-Y subunits (A, B, C) were jointly expressed (Additional file 1: Figure S4B). We also found that NF-Y must not necessarily bind to the DNA to reduce the transactivating potential of C/EBP β . This was shown by replacing three amino acids in the factor NF-YA. This replacement is known to abolish DNA binding of the NF-Y complex [26]. We found that inclusion of this mutated NF-YAm29 subunit was as efficiently quenching the C/EBP β mediated PIA stimulation as the wild type NF-YA subunit (Additional file 1: Figure S4B).

The observation that the NF-Y factors might exert in trans their blocking effect on the C/EBP β mediated activation of PIA was experimentally augmented. We expressed together in HEK293 cells the factors NF-YAm29, NF-YB, NF-YC and C/EBP β and cotransfected the short PIA reporter gene. We analyzed from the same cells in ChIP assays if NF-Y factors may physically interact with C/EBP β having bound to PIA. To this end, we measured in qPCR the amount of promoter molecules recovered by antibodies directed against C/EBP β and NF-YA. We choose PCR amplification primers spanning the C/EBP site, but not the NF-Y binding site. We found that the antibodies against both factors retrieve approximately similar amounts of PIA (Figure 4D). These data show that NF-Y factors can indeed bind in trans to the C/EBP factor.

Different tissue-specific levels of both, PIA activity and C/EBP β mRNA concentrations

We examined if our in vitro derived data concerning mechanism regulating PIA activity are of physiological relevance in vivo. We exploited for this purpose tissue samples derived from two different stacks of animals. Livers were taken from those 12 lactating cows having

been fed normal and energy reduced straw diets, having above already been referred to. In addition, adipose and mammary gland tissues were collected from 16 mid lactating cows having been fed 30 days before slaughter either a norm diet or a fat enriched diet. Experimental conditions for those two groups of animals have extensively been described [28,30].

We compared the PIA activity in different tissues to the mRNA abundance of both families of transcription factors. The concentration of PIA derived transcripts differed >100-fold between liver and lactating mammary gland, while intermediate concentrations were found in adipose tissue (Table 1). This indicates substantial tissue-specific differences in PIA activity. The concentration of NF-YA-encoding mRNA molecules was fairly constant. However, the concentrations of C/EBP β varied between the tissues. Analysis of a potential interdependence between the mRNA concentrations of those factors revealed a strikingly clear correlation of PIA derived transcripts with the ratio between the tissue-specific mean values of NF-YA mRNA copies over those of C/EBP β (Additional file 1: Figure S5). The correlation between these parameters is statistically high and significant (full symbols; r , 0.997; $p < 0.001$). Moreover, including the mean values derived from the different nutritional subgroups (Table 1) into this correlation analysis reveals that the nutrition-dependent different mean values also fit very well to this correlation (open symbols, Additional file 1: Figure S5). However, due to the eventually large individual variation, not all differences between the subgroup means of a given tissue are statistically significant cf (Table 1). No significant degree of correlation was found if all the individual data were considered, due to the high degree of individual variation. We found the highest degree of individual variability for the levels of PIA derived transcripts (CV, 116), while the same parameter was much lower for the mRNA concentrations of those transcription factors (52% and 5%, for C/EBP β and NF-YA respectively).

The mRNA concentrations of both factors are very different in the respective model cells. We found in those pbMEC cultures from cows ~40fold higher concentrations of the NF-YA ($3.1 \pm 0.2 \times 10^5$, $n = 3$) and only ~1% of the C/EBP β mRNA concentration ($5.4 \pm 0.32 \times 10^3$, $n = 3$) than found in vivo in udder tissue. This very low endogenous concentration of the C/EBP β mRNA in the pbMEC model cells may relate to the observation that overexpression of the C/EBP β mRNA encoding vector in these cells significantly increased the activity of the long PIA-reporter construct. This would not be predicted by the above model. The C/EBP β mRNA

concentration in pbMEC is very similar to that previously reported from us for the murine HC-11 MEC model cell [11]. Neither of these model cells endogenously expresses PIA.

Discussion

We initiated this study in order to characterize the molecular principle repressing the general activity of PIA of the ACC- α -encoding gene in cattle. To ensure the physiological relevance of our analysis, we first validated for the cow that PIA, indeed is the nutritionally regulated promoter. This was previously only assumed to be the case [7] by analogy with the situation found in other animals including rat [31,32]. We also excluded that a more upstream located ACC- α promoter "PI" (EMBL Acc No FN185962) would override those transcriptional controls of PIA. We found this promoter to be prominently expressed in the brain (Shi, unpublished), just as that from sheep [10]. Significantly, transcripts derived from PI do not contain exon1A derived sequences, as revealed by 5'-RACE experiments (EMBL acc No FN185963). Thus, transcripts containing exon1A truly stem from PIA and thus are diagnostic for the activity of PIA.

NF-Y and C/EBP factors may either repress or activate PIA activity, depending on the binding site in the promoter.

Our mapping and mutation data indicate that structurally the bipartite PIA repressor is composed of attachment sites for CCAAT-box binding transcription factors of the NF-Y and of the C/EBP families, binding to the distal promoter. Their repressive strength is additive, at least. However, we also observed that attachment sites for both transcription factors on the proximal promoter function as activators for PIA activity. Hence, the binding position rather than the nature of the CCAAT-box binding factor itself determines its function as either to repress or to activate PIA activity.

The bipartite structure of the PIA repressor in cattle is similar to that of the rat PIA promoter [17]. The 5'-part of the rat PIA repressor is formed by a microsatellite, a CA28 repeat. It is not known which factor might bind there. The distal element of the bovine PIA repressor resides in an Art2 retroposon element [7]. We now found that NF-Y factors may bind there to repress PIA activity. The 3'-part of the repressor was found in rat [16,17] as well as in cattle (this study) to depend on a C/EBP factor binding site. Hence, the structurally divergent, yet physiologically equivalent PIA promoters of rat and cow feature a similar bipartite structure of that repressor. However, the structures of the 5'-segment of that repressor are completely unrelated between cattle and rat.

NF-Y is known to regulate the transcription of many genes by interacting with the CCAATbox sequence motif, either enhancing [22,33,34] or dampening expression [27,35]. Dual NFY binding sites of high and low binding affinities have been reported to reside in the promoter of the human von Willebrand factor (VWF) gene [36]. The low affinity site, located in exon1, was found to repress VWF promoter activity, while the high affinity site functions as an activator, quite similar to our findings in the cattle PIA promoter. Interestingly, the high affinity activator binding site of the human VWF gene spans the positions from -30 to +1, very comparable as found in the bovine PIA. The repressive function of NF-Y involves interaction with histone deacetylase (HDAC). NF-Y recruits HDACs to the VWF promoter thus inhibiting the promoter activity in non-endothelial cells. This repression depends on NFY-binding to that downstream repressive site [37].

Likewise, we found that two C/EBP binding sites are crucially involved in regulating the PIA promoter activity. The proximal binding site contributes to basal promoter activity. Stimulation of gene expression through C/EBP factors has often been found [38-40]. It has also been observed, that C/EBP factors may repress transcription [15,41], eventually through repressive family members, such as C/EBP γ and - ζ , or repressive isoforms [19] or through the interaction with HDAC in various physiology conditions [42]. Hence, also the C/EBP family of transcription factors is known to either positively or negatively regulate gene expression.

Our data show that two criteria determine whether these factors activate or repress PIA promoter activity: First, their binding to either the distal repressor or proximal driver region matters but also, second their eventual interaction.

NF-Y represses the activating capability of C/EBP β via protein-protein interaction

We observe, as a novel mechanism that the functional NF-Y factor complex reduces the strong enhancing effect of C/EBP β by protein-protein interaction. The repressive effect of NF-Y on the C/EBP β mediated activation may be unique to PIA of the ACC- α in cattle. In contrast to our observation, it was found in mouse that the interaction between NF-Y with C/EBP α strongly enhances the stimulatory effect on the amelogenin gene promoter activity. ChIP experiments indicated in this example that NF-Y interacts in trans with C/EBP α [23]. C/EBP δ could eventually substitute for the role of the C/EBP α factor [43]. Similarly, expression of the human microsomal-epoxide-hydrolase gene (EPHX) is stimulated through interaction of promoter bound NF-Y factors with C/EBP α . This interaction must be

mediated in trans, since this promoter does not bind directly the factor C/EBP α [44].

We also found that protein-protein interaction mediates the repressive function of NF-Y factors on the stimulating effect of C/EBP β , based on two lines of evidence. Reporter gene assays involving the mutated DNA-binding incompetent factor NF-YA_{m29} and ChIP data congruently support the conclusion that the repressive effect of NF-Y on C/EBP β mediated enhancement of PIA activity is mediated in trans.

Our data may therefore be summarized in a model of a master control unit regulating PIA activity (Figure 4E). Small amounts of NF-Y and C/EBP β factors are sufficient to saturate the proximal binding sites and to activate the promoter. Hence, their high binding affinity for those factors ensures that PIA in principal could actively transcribe the gene, almost in any tissue given that both transcription factors are ubiquitously present. Indeed, a pivotal role of C/EBP β in lipid synthesis has recently been proven by showing that functional disruption of the C/EBP β -encoding gene results in impaired fat accumulation in adipose tissue [41].

However, recruitment of a transcription factor to a high affinity site cannot reflect different factor concentrations, since high affinity sites respond similarly to a switch (e.g. either “on” or “off”) rather than like an adjustable lever. To nevertheless achieve regulation, PIA activity is thus dampened by the factors binding to those distal, low affinity sites. Much higher factor concentrations are necessary to saturate them. Hence, the probability that they are occupied can reflect different factor concentrations. The very large difference observed between the binding affinities of the proximal vs distal C/EBP binding sites suggests that these might be of pivotal significance in this regard. Once those distal, repressive sites are occupied, they might physically be brought into contact with the reciprocal factor residing on the proximal promoter to repress via protein-protein interaction the activation potential. This would structurally require that the promoter elements separating the different components of the master control unit are looped out.

C/EBP β may be the prominent factor to constrain the tissue-specific PIA activity

PIA activity, as assessed on the basis of PIA derived transcripts differs significantly between tissues, similar as it is found for the balance between the concentrations of NF-YA and C/EBP β -encoding mRNA molecules. The latter ratio is mainly influenced by large tissue-specific differences of the C/EBP β mRNA concentration. Assuming that the mRNA concentrations reflect the factor concentrations we then observe, in keeping with our model

the highest PIA activity in the lactating mammary glands, where also the C/EBP β mRNA concentrations (e.g. factor concentrations) are least (Table 1). Moreover, the diet dependent changes of PIA activity in this gland are inversely correlated with the C/EBP β mRNA concentrations. The average PIA activity is highest in the mammary gland of cows having been fed with a fat-enriched diet and concomitantly we found that the C/EBP β mRNA concentrations are least under these conditions. C/EBP β is also found to be regulated in the mammary gland of the mouse by the fat content of the diet [29], as well as with different stages of lactation [45].

Taken together our data suggest that tissue-specific alterations in the proportion of NF-Y and C/EBP β adjust very different set points for the gross PIA activity around which other factors may finely tune the individual PIA activity according to the acute actual needs.

Model cells do not properly reflect the situation in the tissues

It would have been desirable to validate this model through manipulation of the ratio of NF-Y / C/EBP β factors. However, the initial mRNA concentrations for both factors are so vastly different in the pbMEC cells compared to the situation found in the udder tissue that a direct comparison is not really meaningful. Indeed, it is long known that these pbMEC express neither genes for milk protein- nor milk fat-synthesis [46]. They also do not express the endogenous ACC- α PIA (own measurements, data not shown). Moreover, we previously attempted to establish primary cultures of liver cells from cattle as relevant cell model for the analysis of PIA regulation. We found that within 1 h after extirpation the cells would only feature 10% of the original PIA mRNA concentration and another 2.5 h later this concentration was barely measurable, down to ~3% of the original mRNA concentration.

Hence, these cell models do not maintain and reflect the developmentally finely tuned situation found in vivo in the specific tissue within the organ.

Conclusions

The proximal promoter segment of PIA appears to be principally in an active state, since even minute concentrations of both, NF-Y and C/EBP β factors can saturate the high affinity activator sites. Higher factor concentrations will saturate the low affinity repressive sites on the distal promoter resulting in reduced and calibrated promoter activity. Based on measurements of the mRNA concentrations of those factors in different tissues we propose that the interplay of both factors may set tissue-specific limits for PIA activity.

Methods

Animals

All animal experiments were performed according to protocols approved by the relevant authorities (Landesamt für Landwirtschaft, Lebensmittelsicherheit und Fischerei, Mecklenburg-Vorpommern).

Reporter gene constructs

All oligonucleotides used throughout the study are listed in Additional file 1: Table S1. Establishment of the constructs in pGL3basic (Promega) is described in Additional file 2: Supplementary materials and methods.

Fusion PCR

Fusion PCR was performed as described [47]. All primers are listed in Additional file 1: Table S2, and a detailed protocol is given in Additional file 2: Supplementary method.

Cell culture, transient transfection and luciferase assays

Cells from the established murine mammary epithelial cell line (HC-11), primary bovine Mammary Epithelial Cells (pbMEC) and human Embryonic Kidney Cells (HEK293) were cultured as previously described [7,48]. Transient transfection and luciferase assays were done essentially as described [7]. Co-transfection of the vector pRL-TK renilla (Promega) allowed to eventually normalizing for slightly varying transfection efficiencies. The normalized data were expressed as multiples of the activity of the promoterless pGL3basic clone, the hosting vector for the different promoter fragments.

Quantitative real-time PCR

The abundances of ACC- α transcripts derived from PIA, PII, or from all promoters (total), and those encoding NF-YA and C/EBP β were measured by quantitative Real-time PCR (RT-qPCR) assays as described [7,8]. Sampling of the biopsies from liver, mammary gland and fat was already described [28]. Tissues were snap-frozen in liquid nitrogen immediately after collection. One hundred milligram of ground tissue in 1 ml of TRIzol reagent (Invitrogen) was homogenized with Ultra-Turrax T25 for 10 s. After addition of 0.2 ml of chloroform and centrifugation at 12,000 g for 15 min at 4°C, aqueous phase was transferred for total RNA precipitation by adding 1 volume of isopropanol. The RNA pellet was washed with 1 ml of 70% ethanol and air-dried. RNA was dissolved in DEPC-treated water and stored at -70°C. RNA quality was analyzed by agarose gel electrophoresis containing formaldehyde and RNA concentration was measured with a Nanodrop spectrophotometer. Two μ g of total RNA was first primed in reverse with oligo (dT)₂₀ and a gene specific primer (primers listed in Additional file 1: Table S3) using Superscript II

reverse transcriptase (Invitrogen) as recommended by supplier. The first-strand cDNA was purified with the High Pure PCR Product Purification Kit (Roche) and eluted with 100 μ l of water. Real-time PCR was carried out with the LightCyclerFastStart DNA Master SYBR Green I Kit (Roche). The 10 μ l assays contained 5 μ l of cDNA template, 2.6 μ l of water, 0.2 μ l of each 25 μ M forward and reverse primers and 2 μ l of the master mix. Pre-incubation was performed at 95°C for 10 min, and amplification was carried out as 95°C 15s; 60°C 5s; 72°C 20s for 40 cycles. Primers and amplicon sizes are described in Additional file 1: Table S3. Beta-actin was used to normalize transcription data. A dilution series (10¹-10⁶copies) of the appropriate cDNA subclones were included in each run and served as an external standard to calculate the copy numbers of transcripts. All assays were run in duplicate.

Electrophoretic mobility shift assay

Preparation of nuclear extracts and Electrophoretic Mobility Shift Assays (EMSA) were performed as described [48]. Probes were labeled radioactively with ³²P-dCTP using the Klenow-fill in procedure, as described [9]. To this end, we primed the fill-in reaction on the respective "long" oligonucleotide (Additional file 1: Table S4) by annealing the corresponding "short" reverse oligonucleotide. Labeled probes and nuclear extracts were incubated with 10 μ l of 2x binding buffer (20 mM HEPES pH7.9, 10 mM MgCl₂, 0.2 mM EDTA pH8, 2 mM DTT, 0.1 M NaCl, 20% Glycerol, 0.4 mM PMSF) in 20 μ l of volume. DNA-protein complexes were separated on 4% or 6% (NF-Y or C/EBP, respectively) non-denaturing polyacrylamide gels. Anti-NF-YA (sc-10779x) and anti-C/EBP β (sc-746x) antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA).

Chromatin Immunoprecipitation assays

The general procedures for chromatin immunoprecipitation assays (ChIP) were essentially as described [49]. Details of the experiment are shown in Additional file 2: Supplementary methods.

Statistical analysis

The statistical significance of differing mean values was assessed with Student's t-test, as provided with the Excel program by the Microsoft Office software.

Additional files

Additional file 1: Supplementary Figures and Tables.

Additional file 2: Supplementary Material and methods. [7,47,50-52].

Abbreviations

ACC: α (eg greek alpha)acetyl-CoA carboxylase-alpha; PIA: promoter IA; C/EBP: CCAAT/enhancer binding protein; NF-Y: Nuclear factor-Y; EMSA: Electrophoretic mobility shift assay; qPCR: Quantitative Real-time PCR; ChIP: Chromatin immunoprecipitation; HC-11: Murine mammary epithelial cells; HEK293: Human embryonic kidney cells.

Competing interests

The authors declare that there are not any financial or non-financial competing interests associated with this work.

Authors' contributions

XS conducted the molecular biology experiments, has contributed developing the design of the study and drafted the ms. CM contributed to designing the study and the animal experimentation. HMS initiated and supervised the work and finalized the manuscript. All authors read and approved the final manuscript.

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References

- Kim KH: Regulation of mammalian acetyl-coenzyme A carboxylase. *Annu Rev Nutr* 1997, **17**:77-99.
- Mao J, Chirala SS, Wakil SJ: Human acetyl-CoA carboxylase 1 gene: Presence of three promoters and heterogeneity at the 5'-untranslated mRNA region. *Proc Natl Acad Sci USA* 2003, **100**:7515-7520.
- Brusselmans K, De Schrijver E, Verhoeven G, Swinnen JV: RNA Interference-Mediated Silencing of the Acetyl-CoA-Carboxylase- α Gene Induces Growth Inhibition and Apoptosis of Prostate Cancer Cells. *Cancer Res* 2005, **65**:6719-6725.
- Wakil SJ, Abu-Elheiga LA: Fatty acid metabolism: target for metabolic syndrome. *J Lipid Res* 2009, **50**:S138-S143.
- Loor JJ, Everts RE, Bionaz M, Dann HM, Morin DE, Oliveira R, et al: Nutrition-induced ketosis alters metabolic and signaling gene networks in liver of periparturient dairy cows. *Physiol Genomics* 2007, **32**:105-116.
- Barber MC, Price NT, Travers MT: Structure and regulation of acetyl-CoA carboxylase genes of metazoa. *Biochim Biophys Acta* 2005, **1733**:1-28.
- Mao J, Marcos S, Davis SK, Burzlaff J, Seyfert HM: Genomic distribution of three promoters of the bovine gene encoding acetyl-CoA carboxylase alpha and evidence that the nutritionally regulated promoter I contains a repressive element different from that in rat. *Biochem J* 2001, **358**:127-135.
- Mao J, Seyfert HM: Promoter II of the bovine acetyl-coenzyme A carboxylase-alpha-encoding gene is widely expressed and strongly active in different cells. *Biochim Biophys Acta* 2002, **1576**:324-329.
- Mao J, Molenaar AJ, Wheeler TT, Seyfert HM: STAT5 binding contributes to lactational stimulation of promoter III expressing the bovine acetyl-CoA carboxylase alpha-encoding gene in the mammary gland. *J Mol Endocrinol* 2002, **29**:73-88.
- Travers MT, Cambot M, Kennedy HT, Lenoir GM, Barber MC, Joulin V: Asymmetric expression of transcripts derived from the shared promoter between the divergently oriented ACACA and TADA2L genes. *Genomics* 2005, **85**:71-84.
- Shi X, Liu S, Metges CC, Seyfert HM: C/EBP-beta drives expression of the nutritionally regulated promoter IA of the acetyl-CoA carboxylase-alpha gene in cattle. *Biochim Biophys Acta* 2010, **1799**:561-567.
- Molenaar A, Mao J, Oden K, Seyfert HM: All Three Promoters of the Acetyl-Coenzyme A-Carboxylase [alpha]-encoding Gene Are Expressed in Mammary Epithelial Cells of Ruminants. *J Histochem Cytochem* 2003, **51**:1073-1081.
- Lopez-Casillas F, Kim KH: The 5' untranslated regions of acetyl-coenzyme A carboxylase mRNA provide specific translational control in vitro. *Eur J Biochem* 1991, **201**:119-127.
- Travers MT, Vallance AJ, Gourlay HT, Gill CA, Klein I, Bottema CB, et al: Promoter I of the ovine acetyl-CoA carboxylase-alpha gene: an E-box motif at -114 in the proximal promoter binds upstream stimulatory factor (USF)-1 and USF-2 and acts as an insulin-response sequence in differentiating adipocytes. *Biochem J* 2001, **359**:273-284.
- O'Callaghan BL, Koo SH, Wu Y, Freaque HC, Towle HC: Glucose Regulation of the Acetyl-CoA Carboxylase Promoter PI in Rat Hepatocytes. *J Biol Chem* 2001, **276**:16033-16039.
- Tae HJ, Zhang S, Kim KH: cAMP Activation of CAAT Enhancer-binding Protein-beta Gene Expression and Promoter I of Acetyl-CoA Carboxylase. *J Biol Chem* 1995, **270**:21487-21494.
- Tae HJ, Luo X, Kim KH: Roles of CCAAT/enhancer-binding protein and its binding site on repression and derepression of acetyl-CoA carboxylase gene. *J Biol Chem* 1994, **269**:10475-10484.
- McKnight SL, Lane MD, Gluecksohnwaelsch S: Is Ccaat/Enhancer-Binding Protein A Central Regulator of Energy-Metabolism. *Genes Dev* 1989, **3**:2021-2024.
- Schrem H, Klempnauer J, Borlak J: Liver-Enriched Transcription Factors in Liver Function and Development. Part II: the C/EBPs and D Site-Binding Protein in Cell Cycle Control, Carcinogenesis, Circadian Gene Regulation, Liver Regeneration, Apoptosis, and Liver-Specific Gene Regulation. *Pharmacol Rev* 2004, **56**:291-330.
- Descombes P, Schibler U: A Liver-Enriched Transcriptional Activator Protein, Lap, and A Transcriptional Inhibitory Protein, Lip, Are Translated from the Same Messenger-Rna. *Cell* 1991, **67**:569-579.
- Descombes P, Chojkier M, Lichtsteiner S, Falvey E, Schibler U: Lap, A Novel Member of the C/Ebp Gene Family, Encodes A Liver-Enriched Transcriptional Activator Protein. *Genes Dev* 1990, **4**:1541-1551.
- Park SK, Oh SY, Lee MY, Yoon S, Kim KS, Kim JW: CCAAT/Enhancer Binding Protein and Nuclear Factor-Y Regulate Adiponectin Gene Expression in Adipose Tissue. *Diabetes* 2004, **53**:2757-2766.
- Xu Y, Zhou YL, Luo W, Zhu QS, Levy D, MacDougall OA, et al: NF-Y and CCAAT/Enhancer-binding Protein α Synergistically Activate the Mouse Amelogenin Gene. *J Biol Chem* 2006, **281**:16090-16098.
- Tomita T, Kimura S: Regulation of mouse Scgb3a1 gene expression by NF-Y and association of CpG methylation with its tissue-specific expression. *BMC Mol Biol* 2008, **9**:5.
- Gurtner A, Manni I, Fuschi P, Mantovani R, Guadagni F, Sacchi A, et al: Requirement for down-regulation of the CCAAT-binding activity of the NF-Y transcription factor during skeletal muscle differentiation. *Mol Biol Cell* 2003, **14**:2706-2715.
- Mantovani R, Li XY, Pessara U, Hooft van Huisdijnen R, Benoist C, Mathis D: Dominant negative analogs of NF-YA. *J Biol Chem* 1994, **269**:20340-20346.
- Bernadt CT, Nowling T, Wiebe MS, Rizzino A: NF-Y behaves as a bifunctional transcription factor that can stimulate or repress the FGF-4 promoter in an enhancer-dependent manner. *Gene Expr* 2005, **12**:193-212.
- Kuhla B, Albrecht D, Kuhla S, Metges CC: Proteome analysis of fatty liver in feed-deprived dairy cows reveals interaction of fuel sensing, calcium, fatty acid, and glycogen metabolism. *Physiol Genomics* 2009, **37**:88-98.
- Rudolph MC, McManaman JL, Phang T, Russell T, Kominsky DJ, Serkova NJ, et al: Metabolic regulation in the lactating mammary gland: a lipid synthesizing machine. *Physiol Genomics* 2007, **28**:323-336.
- Duske K, Hammon HM, Langhof AK, Bellmann O, Losand B, Nurnberg K, et al: Metabolism and lactation performance in dairy cows fed a diet containing rumen-protected fat during the last twelve weeks of gestation. *J Dairy Sci* 2009, **92**:1670-1684.
- Lopezcasillas F, Poncecastaneda MV, Kim KH: In vivo Regulation of the Activity of the 2 Promoters of the Rat Acetyl Coenzyme-A Carboxylase Gene. *Endocrinology* 1991, **129**:1049-1058.
- Travers MT, Barber MC: Tissue-specific control of the acetyl-CoA carboxylase gene. *Biochemical Soc Tran* 1997, **25**:1215-1219.
- Borghini S, Vargiolu M, Di Duca M, Ravazzolo R, Ceccherini I: Nuclear Factor Y Drives Basal Transcription of the Human TLX3, a Gene

- Overexpressed in T-Cell Acute Lymphocytic Leukemia. *Mol Cancer Res* 2006, **4**:635–643.
34. Matsuo N, Yu-Hua W, Sumiyoshi H, Sakata-Takatani K, Nagato H, Sakai K, et al: The Transcription Factor CCAAT-binding Factor CBF/NF-Y Regulates the Proximal Promoter Activity in the Human α 1(XI) Collagen Gene (COL11A1). *J Biol Chem* 2003, **278**:32763–32770.
 35. Manni I, Mazza G, Gurtner A, Mantovani R, Haugwitz U, Krause K, et al: NF-Y Mediates the Transcriptional Inhibition of the cyclin B1, cyclin B2, and cdc25C Promoters upon Induced G2 Arrest. *J Biol Chem* 2001, **276**:5570–5576.
 36. Peng YW, Jahroudi N: The NFY transcription factor functions as a repressor and activator of the von Willebrand factor promoter. *Blood* 2002, **99**:2408–2417.
 37. Peng Y, Stewart D, Li W, Hawkins M, Kulak S, Ballermann B, et al: Irradiation modulates association of NF-Y with histone-modifying cofactors PCAF and HDAC. *Oncogene* 2007, **26**:7576–7583.
 38. Reddy KV, Serio KJ, Hodulik CR, Bigby TD: 5-lipoxygenase-activating protein gene expression - Key role of CCAAT/enhancer-binding proteins (C/EBP) in constitutive and tumor necrosis factor (TNF) alpha-induced expression in THP-1 cells. *J Biol Chem* 2003, **278**:13810–13818.
 39. Zhang F, Lin M, Abidi P, Thiel G, Liu J: Specific Interaction of Egr1 and c/EBP β Leads to the Transcriptional Activation of the Human Low Density Lipoprotein Receptor Gene. *J Biol Chem* 2003, **278**:44246–44254.
 40. Zhou YL, Snead ML: Identification of CCAAT/Enhancer-binding Protein alpha as a Transactivator of the Mouse Amelogenin Gene. *J Biol Chem* 2000, **275**:12273–12280.
 41. Schroeder-Gloeckler JM, Rahman SM, Janssen RC, Qiao L, Shao J, Roper M, et al: CCAAT/Enhancer-binding Protein beta Deletion Reduces Adiposity, Hepatic Steatosis, and Diabetes in Leprdb/db Mice. *J Biol Chem* 2007, **282**:15717–15729.
 42. Wang GL, Salisbury E, Shi XR, Timchenko L, Medrano EE, Timchenko NA: HDAC1 cooperates with C/EBP alpha in the inhibition of liver proliferation in old mice. *J Biol Chem* 2008, **283**:26169–26178.
 43. Xu Y, Zhou YL, Gonzalez FJ, Snead ML: CCAAT/enhancer-binding protein delta (C/EBPdelta) maintains amelogenin expression in the absence of C/EBPalpha in vivo. *J Biol Chem* 2007, **282**:29882–29889.
 44. Zhu QS, Qian B, Levy D: CAAT/Enhancer-binding Protein α (C/EBP α) Activates Transcription of the Human Microsomal Epoxide Hydrolase Gene (EPHX1) through the Interaction with DNA-bound NF-Y. *J Biol Chem* 2004, **279**:29902–29910.
 45. Gigliotti AP, DeWille JW: Lactation status influences expression of CCAAT/enhancer binding protein isoform mRNA in the mouse mammary gland. *J Cell Physiol* 1998, **174**:232–239.
 46. Günther J, Koczan D, Yang W, Nürnberg G, Repsilber D, Schuberth HJ, Park Z, Maqbool N, Molenaar A, Seyfert HM: Assessment of the immune capacity of mammary epithelial cells: comparison with mammary tissue after challenge with *Escherichia coli*. *Vet Res* 2009, **40**:31.
 47. Liu S, Shi X, Bauer I, Gunther J, Seyfert HM: Lingual antimicrobial peptide and IL-8 expression are oppositely regulated by the antagonistic effects of NF-kappaB p65 and C/EBPbeta in mammary epithelial cells. *Mol Immunol* 2011, **48**:895–908.
 48. Yang W, Molenaar A, Kurts-Ebert B, Seyfert HM: NF-kappa B factors are essential, but not the switch, for pathogen-related induction of the bovine beta-defensin 5-encoding gene in mammary epithelial cells. *Mol Immunol* 2006, **43**:210–225.
 49. Zhan X, Shi X, Zhang Z, Chen Y, Wu JI: Dual role of Brg chromatin remodeling factor in Sonic hedgehog signaling during neural development. *Proc Natl Acad Sci USA* 2011, **108**:12758–12763.
 50. Nowak DE, Tian B, Brasier AR: Two-step cross-linking method for identification of NF-kappa B gene network by chromatin immunoprecipitation. *Biotechniques* 2005, **39**:715–725.
 51. Zhao JQ, Glasspool RM, Hoare SF, Bilisland A, Szatmari I, Keith WN: Activation of telomerase rna gene promoter activity by NF-Y, Sp1, and the retinoblastoma protein and repression by Sp3. *Neoplasia* 2000, **2**:531–539.
 52. Wu J, Metz C, Xu X, Abe R, Gibson AW, Edberg JC, Cooke J, Xie F, Cooper GS, Kimberly RP: A Novel Polymorphic CAAT/Enhancer-Binding Protein beta Element in the FasL Gene Promoter Alters Fas Ligand Expression: A Candidate Background Gene in African American Systemic Lupus Erythematosus Patients. *J Immunol* 2003, **170**:132–138.

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